ANTIFUNGAL SESQUITERPENOIDS FROM AN ARTHROCONIDIAL FUNGUS

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ABSTRACT.—The metabolites produced in liquid culture by an arthroconidial fungus have been examined, and six new hirsutane metabolites have been isolated from the antifungal neutral extract. The structures of the major metabolites, which we have named arthrosporone [1], anhydroarthrosporone [2], and arthrosporol [3], have been determined by chemical and spectroscopic methods.

As part of a continuing search for naturally occurring antifungal agents we have investigated the metabolites produced by an arthroconidial fungus (UAMH 4262) when grown in liquid still culture. The as yet unidentified fungus was encountered accidentally by Tsuneda and Hiratsuka as a contaminant on a culture of the phytopathogenic fungus *Ceratocystis ulmi* (Buis.) Moreau, the causative agent of Dutch elm disease (1,2). The fungus has been examined by Dr. J. Stalpers who considered it to be a haploid basidiomycete, probably belonging to a wood-inhabiting Agaricales. The sexual stage is unknown, and in culture the fungus produces an asexual stage consisting of rectangular arthroconidia with short separating cells, similar to those described as Group 3 (3). No hyphomycete genus is currently available to accommodate these basidiomycete anamorphs¹. In competitive culture the arthroconidial fungus strongly inhibits *C. ulmi* and other wood-destroying fungi of the genus *Ceratocystis*, including those that are responsible for the blue stain disease of pine (4–6).

Fungus UAMH 4262 was grown on a medium of potato dextrose broth in still culture for approximately 30 days. Bioassays of the culture extracts revealed that the antifungal activity was located mainly in the neutral fraction of the broth extracts (7). The neutral fraction was separated into several components by Si gel chromatography. These components were further purified by Si gel chromatography and, in some instances, fractional crystallization. This led to the isolation of several new sesquiterpenoids as well as the known sesquiterpene tricyclohumuladiol (8–10). We report herein the structures of these new sesquiterpenes.

RESULTS AND DISCUSSION

In order to facilitate the discussion, the chemistry of the arthroconidial metabolites that led to the derivation of the complete structures of these previously unknown compounds is presented in terms of the final structure of arthrosporone [1] and the closely related anhydroarthrosporone [2]. Arthrosporone ($C_{15}H_{24}O_3$, established by hrms and cims) and anhydroarthrosporone ($C_{15}H_{22}O_2$) are the most abundant crystalline metabolites and are optically active. The ir spectrum of 1 shows hydroxyl (3440 cm⁻¹), carbonyl (1731 cm⁻¹), and gem-dimethyl (1381, 1360 cm⁻¹) absorption bands. The ketonic nature of the carbonyl is apparent from its uv (λ max 280 nm) and ¹³C-nmr spectra (δ 216.4) and by the fact that arthrosporone [1] is readily reduced with NaBH₄ to arthrosporol [3] and its epimer 4. Both the ir and the ¹³C-nmr spectra suggest a cyclopentanone carbonyl.

The tricyclic nature of arthrosporone [1] is apparent from examination of its 13 C-nmr spectrum which shows the ketonic carbonyl carbon and 14 sp³ carbons: four

¹L. Sigler, University of Alberta Microfungus Collection, personal communication, June 1989.

quaternary carbons (two of which bear an oxygen), two CH, four CH₂, and four Me groups (11). The ¹³C- and ¹H-nmr spectra account for 22 hydrogens which are directly attached to carbon; thus, the remaining two hydrogens must be present as part of hydroxyl groups. The hydroxyl groups of arthrosporone [1] are tertiary hydroxyl groups, as revealed by examination of the ¹H-nmr spectrum. There are no lowfield hydrogens (i.e., $\delta > 3.0$), indicating that the molecule contains no olefinic hydrogens or hydrogens geminal to an oxygen atom. The remainder of the spectrum consists of three methyl singlets, a methyl doublet, and several well-resolved one-hydrogen spin-multiplet systems (Table 1). The lack of reactivity of $\mathbf{1}$ in oxidation reactions is consistent with the tertiary nature of its hydroxyl groups. Under forcing basic conditions (Ac₂O, 4-N, Ndimethylaminopyridine, NEt₃, 3 days), arthrosporone is converted to a monoacetyl derivative 5, while acid-catalyzed acetylation provides the diacetyl derivative 6. The ir spectrum of 5 shows hydroxyl (3449 cm⁻¹), carbonyl, and acetoxyl (1736, 1249 cm⁻¹) absorption bands. Its ¹H-nmr displays acetyl methyl (δ 1.98, s) and hydroxyl hydrogen (δ 1.47, s, D₂O exchangeable). This information confirms the presence of two hydroxyl groups in arthrosporone. One hydroxyl group in arthrosporone [1] forms part of a β -hydroxyketone functionality because 1 is readily dehydrated under acid catalysis (p-TsOH, C_6H_6 , 78°, 3 h) to anhydroarthrosporone [2], an α , β -unsaturated ketone.

Anhydroarthrosporone [2], $C_{15}H_{22}O_2$, has a uv spectrum which shows an unsaturated carbonyl chromophore $[\lambda_{max} 230 \ (\epsilon = 13\ 700)]$, while its ir spectrum shows absorption bands characteristic of a hydroxyl (3462 cm⁻¹), an α,β -unsaturated ketone (1693 and 1632 cm⁻¹), and a *gem*-dimethyl group (1372, 1364 cm⁻¹). The ¹³C-nmr spectrum of anhydroarthrosporone [2] displays three lowfield signals for sp²-hybridized carbons at δ 211.6 (weak singlet, carbonyl carbon), δ 177.0 and δ 122.9 [singlet and doublet; disubstituted and monosubstituted olefinic carbons, respectively (12)]. A doublet at δ 63.4 suggests the presence of a methine carbon α to a carbonyl (13). These data suggest that anhydroarthrosporone contains a β -substituted α,β -unsaturated cyclopentenone. The ¹³C-nmr spectrum also shows the presence of a quaternary carbon bearing an hydroxyl group (δ 92.7)(14,15), a quaternary carbon, two CH, three CH₂, and four Me groups.

The ¹H-nmr spectrum of anhydroarthrosporone [**2**] indicates a vinyl hydrogen (δ 5.85) coupled to a methylene hydrogen (δ 2.72), methyl hydrogens (δ 1.11, d) coupled to a vicinal methine hydrogen (δ 2.34, J = 7 Hz), three quaternary methyl groups [δ 1.20 (s), 1.13 (s), 0.94 (s)] and a series of one-hydrogen multiplets (Table 1). The hrms of anhydroarthrosporone [**2**] exhibits fragment ions at m/z 216 [M – H₂O]⁺, 123 [M – C₇H₁₁O]⁺, and base peak at m/z 122 (C₈H₁₀O).

All the spectroscopic data presented for anhydroarthrosporone and its acetyl derivative, O-acetylanhydroarthrosporne [7], are consistent with either a hirsutane skeleton [2] or with skeleton 8. The ¹H-nmr spectrum of O-acetylanhydroarthrosporone [7] (Table 1), when compared with that of anhydroarthrosporone, shows deshielding of a methine hydrogen, two methylene hydrogens, and one of the quaternary methyl hydrogens. For either structure 2 or 8 the deshielded hydrogens would be those vicinal and cis to the acetoxyl group (16). The carbon skeleton of the arthroconidial metabolites was deduced in the following way. Compounds such as 2 or 8 would be expected to undergo a retro-aldol cleavage to give bicyclic compounds 9 or 10, respectively. Compound 9 may be distinguished from 10 by its ir spectrum [cyclobutanone carbonyl absorption near 1780 cm⁻¹, cyclopentanone carbonyl absorption around 1745 cm⁻¹ (17, 18)]. Accordingly, treatment of anhydroarthrosporone with excess NaH in dry C_6H_6 gave a single product in quantitative yield. The product is less polar than the starting metabolite; it is optically active and has a molecular formula of $C_{15}H_{22}O_2$ (hrms), as expected. Its uv spectrum shows a maximum characteristic of an β -alkyl substituted cyclopentenone [λ_{max} 223 nm (observed), 226 nm (calculated) (19)]. Its ir spectrum does not show hydroxyl absorption but has a strong carbonyl absorption band at 1738 cm⁻¹ indicative of the presence of a cyclopentanone as well as strong absorption bands due to an α , β -unsaturated cyclopentenone. The hrms fragmentation pattern of the product shows a base peak at m/z 123 (C₈H₁₀O + H). These data suggest that the retro-aldol cleavage product has structure **9**. The ¹H-nmr spectrum of **9** (see Experimental) gives further support to the assigned structure.

It follows that anhydroarthrosporone possesses the hirsutane skeleton 2 and that 0acetylanhydroarthrosporone is 7. The assignment of the relative stereochemistry in the arthroconidial metabolites was based on the results of the following experiments. The cis, anti, cis configuration consistently found in the hirsutane series is maintained, and the angular methyl at C-2 is arbitrarily assigned a β orientation. The assignment is supported by ¹H-nmr pyridine shift studies (16, 17, 20) [$\Delta = \delta$ (CDCl₃) – δ (C₅D₅N)] carried out on compound 2, which showed that the C-8 hydroxyl is cis to H-1 ($\Delta = -0.41$) and trans to the angular methyl group at C-2 ($\Delta = -0.08$). Because no conclusions could be drawn about the stereochemistry of the secondary methyl group at C-3, ¹H-nmr pyridine-induced shift studies were also carried out on several derivatives of 2.

Reduction of 0-acetylanhydroarthrosporone [7] with NaBH₄/CeCl₃ (21) in MeOH gave a mixture of the allylic alcohol **11** and the epimeric saturated acetoxyalcohols **12** and **13**. The composition of the reduction product was not affected by reaction temperature, order of addition of reagents, or omission of CeCl₃. The reduction of the C,C



double bond of 0-acetylanhydroarthrosporone [7] is expected to take place from the less sterically hindered β face of the molecule because reduction of the structurally similar bicyclic enone 14 proceeds in this manner (22). A pyridine-induced shift study of 11 shows that H-3 is strongly deshielded ($\Delta = -0.46$) whereas the methyl group at C-2 shows no appreciable shift (Table 1) on solvent change. These observations suggest that

$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	Proton	Solvent	Compound							
H-1 CDC13 2.56.dd (8,12) 2.86.dd (9,11) 2.86.dd (8,12) 2.87.dd (8,12) 2.87.dd (8,13)			1	2	5	6	7	11	12	13
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	H-1	CDCl ₃	2.56 dd	2.38 dd	2.83 dd	2.86 dd	2.68t(10)	2.57 dd	2.67 dd	2.67 dd
pridine-d 2.77 dd 2.63 r (10) 2.97 dd 8.70 r (10) 2.87 dd 2.84 dd 2.84 dd H-3 CDCl ₃ 2.57 q (7) 2.32 q (7) 2.32 q (7) 2.32 q (7) 2.33 q (7) 1.81 dd [8,12) pyridine-d 2.92 dq 2.41 q (7.) 2.52 q (1,7) 2.36 q (7.)		-	(8,12)	(9,11)	(9,12)	(8,12)		(9,10)	(8,13)	(8,12.5)
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$		pyridine-d,	2.77 dd	2.63t(10)	2.97 dd			2.70 t (10)	2.85 dd	2.84 dd
H-3 CDC1 ₃ 2.77 q(7) 2.32 q(7) 2.32 q(7) 2.32 q(7) 2.33 q(7) 1.81 dq 1.97 dq 2.35 q(7) pyridine-4, 2.92 dq 2.41 q(7.5) 2.52 q 1.7 2.26 dq 2.34 dq 2.34 dq H-4 CDC1 ₃ 2.69 dq 5.85 (1.3) 2.64 dd 3.21 dd 1.75 d(2) 2.29 d(2) 4.94 m 4.18 m 4.56 m 4.94 m H-5 CDC1 ₃ 2.69 dd 5.85 (1.3) 2.64 d(1) 1.19 m 2.29 d(2) 2.29 d(1) 2.97 d(2) 2.98 dd 4.94 m 4.18 m 4.56 m pyridine-4, 2.75 d(2) 5.92 d(2) 2.69 d(1) 2.29 d(1) 5.66 d(2) 2.04 dd 3.81 d(1) pyridine-4, 2.75 d(1) 2.79 d(16) 2.81 d(10) 3.11 d(1) 3.02 d(1) 3.02 d(1) 2.40 d(1) <td< td=""><td></td><td></td><td>(9.5,12)</td><td></td><td>(8,12)</td><td></td><td></td><td></td><td>(8,12)</td><td>(8,12.5)</td></td<>			(9.5,12)		(8,12)				(8,12)	(8,12.5)
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	H-3	CDCl,	2.57 q(7)	2.32 q(7)	2.34 dq	2.27 dq	2.33 q (7.5)	1.81 dq	1.97 dq	2.35 m
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H-4 CDCl ₃ (1,7) (1,7) <t< td=""><td></td><td>pyridine-d5</td><td>2.92 dq</td><td>2.41g(7.5)</td><td>2.52 q</td><td></td><td></td><td>2.26 dq</td><td>2.38 dq</td><td>2.34 dq</td></t<>		pyridine-d5	2.92 dq	2.41g(7.5)	2.52 q			2.26 dq	2.38 dq	2.34 dq
H-4 CDCl ₃ μ - μ μ - μ <t< td=""><td></td><td></td><td>(1,7)</td><td></td><td></td><td></td><td></td><td>(6,7)</td><td>(7,8)</td><td>(4,8)</td></t<>			(1,7)					(6,7)	(7,8)	(4,8)
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$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	H-5	CDCl ₃	2.69 dd	5.85 d (1.3)	2.64 dd	3.21 dd	5.75 d (2)	5.29 br s	2.08 m	1.96 ddd
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$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$			2.20 d (20)		2.22 d (19)	2.29 d (19)			1.38 ddd	1.88 m
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$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			2 47 11		2 44 1/10				(4,8,11)	(3,8,14)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			2.4/dd		2.44 0 (19)				1./) ddd	2.01 ddd
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	цζ		(1,20)						(4,0,1)	(4,0,14)
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$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	H_7	CDC1	2 40 4 (16)	2 70 4 (16)	2 81 4 (16)	3 31 4(17)	3 30 4 (17)	3 02 dd	2.40 m	2 37 dd
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	11-7	CDC13	2.40 (10)	2.790(10)	2.010(10)	5.514(1/)	5.504(17)	(1, 17)	(11 18)	(9.10)
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$			2 214(16)	2 72 dd	2 24 d	2 184(17)	2 74 4	2.33	2 08 dd	(7,10)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			2.214(10)	(1.16)	(15.5)		(2.17)	(2.17)	(8.18)	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		pyridine-d-	2.75 d(16)	2.96 d (15)	3.10 d		(_,_,	3.29 d (16)	2.59 dd	2.46 dd
H-9 2.46 d(16) (2,15) 2.80 d(1) (2,15) 2.45 d(16) (2,15) 2.45 d(16) (2,15) 2.46 d(1) (2,15) 2.46 d(1) (2,16) 2.46 d(1) (2,16) 2.16 d(1) (2,16) 2.10 d(1) H-9 CDCl ₃ 1.96 d(14) 1.87 dd 2.28 dd 2.31 d(15) 2.43 d(15) 2.29 dq 2.11 dd 1.80 d(15) 1.79 dd 1.68 d(14) 1.86 d(15) 1.59 d(15) 1.64 d(15) 1.53 d(15) 1.75 d(15) 1.85 d(15) pyridine-d ₂ 2.14 d(14) 2.08 dd 2.46 d(15) 1.59 d(15) 1.64 d(15) 1.59 d(15) 1.55 d(15) 1.85 d(15) 1.85 d(15) (3,14) 2.06 dd 1.68 d(14) 2.15 d(15) 1.67 r(12) 1.69 ddd 1.52 ddd 1.83 d(15) 1.85 d(15) (3,14) 1.70 r(12) 1.72 ddd 1.65 ddd 1.67 r(12) 1.69 ddd 1.52 ddd 1.30 r(13) (3,8,13) 1.59 ddd 1.79 r(12) 1.65 ddd 2.00 r(12) (11,13) (3,8,12) (11,13) (3,8,13) (3,8,13) (3,8,13) (3,8,13) (3,8,13) (3,8,13) (3,8,13		[]]			(15.5)				(9,15)	(9,15)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			2.46d(16)	2.80 dd	2.45 d (16)			2.46 dd	2.46 dd	2.11 dd
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $				(2,15)				(2,16)	(4,8)	(7,15)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	H-9	CDCl,	1.96d(14)	1.87 dd	2.28 dd	2.31d(15)	2.43 d (15)	2.29 dq	2.11 dd	1.80 d(15)
H-11 1.79 dd (2.5,14) 1.68 d (14) (2.5,14) 1.86 d (15) (2.6) 1.59 d (15) (2.6) 1.63 d (15) (2.5) 1.75 d (15) (3.15) 1.75 d (15) (3.15) 1.85 d (15) (3.15) H-11 CDCl ₃ 1.69 d (14) (2.7) 2.06 dd (3.14) 1.68 d (14) (2.9,13) 2.16 d (15) (2.9,13) 1.67 t (12) (2.9,13) 1.67 d (15) (2.9,13) 1.85 d (15) (3.15) 1.83 d (15) (2.9,13) 1.85 d (15) (3.15) H-11 CDCl ₃ 1.70 t (12) 1.72 d dd (2.9,13) 1.69 t (12) (2.9,13) 1.67 t (12) (2.9,13) 1.69 d dd (2.9,12.5) 1.48 d dd (1.45 d dd (2.9,13) 1.30 t (13) (3.8,13) pyridine-d ₅ 1.97 t (12) 1.65 d dd (2.9,13) 1.69 d dd (2.9,13) 1.47 d dd (2.9,13) 1.49 d dd (2.9,13) 1.33 d d (2.9,13) 1.25 t (12) 1.31 t (1.13) H-12 CDCl ₃ 1.16s 1.20s 1.04s 1.03s 0.93 s 1.05 s 1.02 s H-13 CDCl ₃ 1.08s 1.13s 0.89 s 1.06s 1.11s 0.80 s 1.02 s H-14		}	}	(2,14)	(3,15)	1	1	(2,15)	(3,15)	1
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			1.79 dd	1.68 d (14)	1.86 d (15)	1.59 d (15)	1.64 d (15)	1.53d(15)	1.75 d (15)	1.85 d (15)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			(2.5,14)							
$ H-11 CDCl_{3} = \begin{bmatrix} (2,14) \\ 1.68 d (14) \\ (3,14) \\ 1.68 d (14) \\ (2,9,13) \\ 1.59 d d \\ (2,9,13) \\ 1.57 t (12) \\ 1.65 d d \\ (2,9,12.5) \\ (2,9,13) \\ (10,13) \\ (10,13) \\ (2,9,13) \\ 1.66 d d \\ (2,9,12.5) \\ (10,13) \\ (10,13) \\ (2,9,13) \\ 1.47 d d \\ (2,9,13) \\ 1.47 d d \\ (3,9,5,12) \\ (11,13) \\ (2,9,13) \\ 1.47 d d \\ (3,9,5,12) \\ (11,13) \\ (3,9,12) \\ (2,9,13) \\ 1.47 d d \\ (3,9,5,12) \\ (11,13) \\ (3,9,12) \\ (2,9,13) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,13) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,12) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,12) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,12) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,12) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,12) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,13) \\ 1.33 d \\ (11,13) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,13) \\ 1.33 d \\ (11,13) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,13) \\ 1.33 d \\ (11,13) \\ 1.47 d d \\ (3,9,5,12) \\ (10,13) \\ (3,8,13) \\ 1.33 d \\ (11,13) \\ 1.47 d d \\ 1.58 d d \\ 1.62 d d \\ 1.52 d d \\ 1.33 d d \\ 1.25 t (12) \\ 1.33 d \\ 1.25 t (12) \\ 1.31 t \\ (11,13) \\ (11,1$		pyridine-d ₅	2.14 d (14)	2.08 dd	2.46 d (15)			2.52 dd	2.29 dd	2.27 dd
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$			2.06.12	(2, 14)	2 16 1/16			(2,15)	(3, 15)	(3,13)
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$			(2.16)	1.08 d (14)	2.13 (15)			1.02 (1)	1.05 (15)	1.85 4(1)
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	H-11	CDCL	$1.70 \cdot (12)$	1 72 ddd	1 69+(12)	1.67+(12)	1 69 ddd	1 52 ddd	1 48 ddd	1 30+(13)
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	11-11	CDCI3	1.701(12)	(2 9 13)	1.0) ((12)	1.0/ ((12)	(2 9 12 5)	(2913)	(3 8 13)	1.500(15)
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$			1.59 ddd	1.47 dd	1.56 ddd	1.62 dd	1.45 dd	1.32 dd	1.22 t(13)	1.53 ddd
$\begin{array}{c c c c c c c c c c c c c c c c c c c $			(2.5.8, 12)	(11.13)	(3,9,12)	(8,12)	(9.12.5)	(10,13)		(3,8,13)
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		pyridine-da	1.97 t(12)	1.65 ddd	2.00 t (12)			1.46 ddd	1.47 ddd	1.49 ddd
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $				(2,9,13)				(2,9,13)	(3,8,13)	(3,8,13)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			1.47 ddd	1.45 dd	1.58 ddd			1.33 dd	1.25 t (12)	1.31 t
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			(3,9.5,12)	(10,13)	(3,8,12)			(11,13)		
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	H-12	CDCl ₃	1.16 s	1.20 s	1. 04 s	1.03 s	0.93 s	1.03 s	1.05 s	1.02 s
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		pyridine-d5	1.24 s	1.40 s	1.01s			1.06 s	1.09 s	1.08 s
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	H-13	CDCl ₃	1.08 s	1.13 s	0.89 s	1.06 s	1.11s	0.80 s	1.05 s	1.04 s
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	/	pyridine-d,	0.96 s	1.10 s	1.07 s			0.83 s	1.03 s	1.02 s
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	H-14	CDCl ₃	U.84s	0.94s	1.08 s	U.88 s	1.09 s	1.05 s	U.80 s	0.975
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		pyridine-a,	0.94 s	0.865	1.09 s	1.05.1(7)	1 10 1/7 5	1.02 s	0.83 s	1.215
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	ri-1)	CLICI3	1.02 d(/)	1.110(/)	1 11 4(7)	1.03 a(/)	[1.100(7.5)	1.090(7.3)	1.05 a(7)	1 25 4 (9)
Operation Operation 1.903	сосн	CDC1.	1.00 a(/)	1.140(/.)	1986	1 98 6	1 98 5	1.20 (1.3)	2 00 4	2 02 5
OH pyridine-d ₅ 1.63 s 1.95 s 1.93 s 1.88 s 2.01 s OH CDCl ₃ 1.63 s 1.47 s 6.34 d (4) 6.26 d (4) 5.82 d (2)	cours	- Constanting				2.04s	1.70 %			
OH CDCl ₃ 1.63 s 1.47 s 2.23 s pyridine-d ₃ 5.60 s 6.21 s 6.34 d(4) 6.26 d(4) 5.82 d(2)		pyridine-d.			1.95 s			1.93 s	1.88 s	2.01 s
pyridine-d ₅ 5.60 s 6.21 s 6.34 d (4) 6.26 d (4) 5.82 d (2)	ОН	CDCI.		1.63 s	1.47 s					2.23 s
		pyridine-d,	5.60 s	6.21 s	1			6.34 d (4)	6.26d(4)	5.82 d (2)

 TABLE 1.
 ¹H-nmr Data^a and Solvent Studies for Compounds 1, 2, 5–7, 11–13.

"Reported as chemical shift (δ), multiplicity, and coupling constant in Hz (in parentheses).

the hydroxyl at C-4 is cis to H-3 and trans to the C-2 methyl. Thus, in allylic alcohol **11** the C-3 methyl is β while the hydroxyl group is α .

The ir spectra of both acetoxyalcohols 12 and 13 show absorption bands characteristic of hydroxyl and acetoxyl functional groups, while the ¹H-nmr spectra of **12** and 13 each show a carbinyl hydrogen, a methyl doublet, an acetoxyl methyl, and three methyl singlets. A pyridine-induced shift study of acetoxyalcohol 12, which was the major isomer, shows that the C-4 hydroxyl is α oriented since it is cis to H-3 $(\Delta = -0.41)$ and trans to the angular methyl at C-2 ($\Delta = -0.03$). On the other hand, the pyridine-induced shift study of the minor epimer, compound 13, shows that the C-4 hydroxyl is β oriented, because the angular methyl at C-2 and the secondary methyl at C-3 each undergo appreciable deshielding when the solvent is changed from $CDCl_3$ to C₅D₅N whereas the chemical shift of H-3 remains unchanged. Correlation experiments were undertaken to confirm the structure of compound 12. Catalytic hydrogenation of anhydroarthrosporone gave a single ketone, 15, which was subjected to further reduction with NaBH₄. The epimeric mixture of diols obtained from the reduction of compound 15 was separated, and the spectral properties of the major product were identical in all respects with those of the diol obtained by deacetylation of acetoxyalcohol 12 with methanolic Na_2CO_3 . It is interesting to note that pyridine shift studies of compounds 11 and 12 show that in each compound the secondary methyl at C-3 undergoes substantial downfield shifts ($\Delta = -0.17$ and -0.19, respectively). This may be explained on the basis of conformational considerations. Ring C of the triquinane system may adopt an envelope (C_3) conformation with C-3 being the flap of the envelope. Examination of molecular models reveals that in the C₃ conformation the C-4 hydroxyl group lies between the hydrogen and the methyl group at C-3.



The stereochemistry of arthrosporone [1] was assigned as follows. The orientation of the C-3 methyl of arthrosporone should be the same as that in anhydroarthrosporone [2], provided that no isomerization took place during the dehydration reaction. Evidence that the stereochemical integrity is retained is provided by ¹H-nmr data: H-3 resonates at δ 2.57 in arthrosporone [1] and at δ 2.54 in ketone 15, indicating a similar environment for this hydrogen in each compound. Comparison of the ¹H-nmr spectra of 1 and 0-acetylarthrosporone [5] reveals that H-3 shifts upfield (-0.22) upon acetylation of the C-8 hydroxyl, indicating that H-3 and the C-8 hydroxyl are proximate to one another. Results of ¹H-nmr pyridine-induced shift studies of compounds 1 and 5 (Table 1) lend support to the stereochemical assignment at C-3 and C-6. In arthro-

sporone, H-3 is strongly deshielded ($\Delta = -0.35$) and the C-2 methyl (vicinal and cis to the C-6 hydroxyl) shows moderate deshielding ($\Delta = -0.10$), while the C-3 methyl is not shifted when the ¹H-nmr spectrum recorded in C₅D₅N is compared with that recorded in CDCl₃. Similar solvent shifts are observed for 0-acetylarthrosporone [**5**]. Finally, nOe studies (23,24) of compound **1** show a 4% enhancement of the C-2 methyl upon irradiation of the C-3 methyl hydrogens. This data confirms that the C-3 methyl is β oriented in arthrosporone and, it follows, in anhydroarthrosporone.

The stereochemistry of arthrosporol is assigned as depicted in structure 3. Oxidation of $\mathbf{3}$ with pyridinium chlorochromate gave arthrosporone $[\mathbf{1}]$ as the only product. This result indicates that the secondary methyl at C-3 has the same orientation in each of compounds 1 and 3. The α orientation of the C-4 hydroxyl and B orientation of the C-6 hydroxyl were established by analysis of the acetoxyl-induced anisotropic shifts of the hydrogens vicinal to the hydroxyl groups of 3 and analysis of its mono-, di-, and triacetyl derivatives. Each derivative was prepared selectively as follows. 4-O-Acetylarthrosporol [16] was prepared by treatment of 3 with Ac₂O in pyridine for 24 h or by treatment with Ac₂O, a catalytic amount of dimethylaminopyridine, and triethylamine for 3 h. The latter reaction, when allowed to proceed for 3 days, gave a quantitative yield of 4,8-0,0-diacetylarthrosporol [17]. 4,6,8,0,0,0-Triacetylarthrosporol [18] was obtained quantitatively when 3 was allowed to stir with a catalytic amount of *p*-toluenesulfonic acid in Ac₂O for 3 h. Table 2 shows the ¹H-nmr data for **3** and its acetyl derivatives. When there is an acetoxyl group at C-4 (e.g., 16, 17, 18), a strong deshielding of H-3 and H-5 is observed. Further substitution of the C-8 hydroxyl with an acetoxyl group results in a downfield shift of H-1, one of H-7, and one of H-9, while H-3, which is very close in space to the C-8 acetoxyl, is shielded. When all three hydroxyl hydrogens are substituted with acetyl groups, both hydrogens at C-5 are deshielded, as would be expected for a trans relationship between the C-4 and C-6 hydroxyl groups. These data confirm the stereochemistry of arthrosporol to be that shown in **3**. Interestingly, 4-0-acetylarthrosporol [16] was isolated as a minor metabolite from the culture broth of the arthroconidial fungus.

Two minor metabolites were isolated from the culture broth. The less polar of these, which we have named dehydroarthrosporodione, has a molecular formula of $C_{15}H_{20}O_3$. It is optically active and contains a secondary hydroxyl (ir 3450 cm⁻¹; ¹³C nmr 75.1 d), a cyclopentanone (ir 1730 cm⁻¹; ¹³C nmr 218.8 s), a cyclopentenone (ir 1700; 1637 cm⁻¹; ¹³C nmr 205.8 s, 185.0 s, 145.0 s) and four methyl groups (¹H and ¹³C nmr). Its uv spectrum shows a maxima at 242 nm (α , β -substituted cyclopentenone), which increases in intensity but does not shift upon the addition of acid or base; this suggests that the hydroxyl group is not part of the chromophore (17, 18). The ¹H-nmr spectrum of dehydroarthrosporodione shows that the carbinyl hydrogen is allylic (δ 4.5, br s) with a small coupling to allylic methylene hydrogens [δ 2.83 (2.2) Hz), 2.21 (2 Hz)]. Decoupling experiments reveal that the allylic methylene hydrogens are also coupled to a methine hydrogen (δ 2.93, m) which is further coupled to two methylene hydrogens [δ 2.60 (4, 19 Hz), 2.39 (2, 9, 19 Hz)]. The size of the geminal coupling observed for the latter methylene indicates that it is α to a carbonyl. These data are consistent with the assignment of structure 19 for dehydroarthrosporodione but are also not inconsistent with the structure in which the carbonyl at C-11 and the hydroxyl at C-9 are interchanged.

Compound 19 readily forms the acetyl derivative 20 and when oxidized with pyridinium chlorochromate gives triketone 21. The spectral properties of compounds 20 and 21 provide support for the substitution pattern in 19. A downfield shift ($\Delta = 0.27, 0.30$) is observed for the allylic methylene hydrogens when the ¹H nmr of 21 is compared with the ¹H nmr of 19. Lack of material precluded further experiments with

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Proton	Compound							
	3	4	16	17	18			
H-1	2.40 dd	2.51 dd	2.41 dd	2.70 dd	2.75 dd			
	(8,12)	(5,14)	(8,11)	(9,12)	(9,12)			
H-3	2.01 m	1.87 d (14)	2.27 dq	2.03 m	2.02 m			
H-4	3.94 dt	4.14 dt	4.84 dt	4.82 ddd	4.85 dt			
	(5,9)	(2,5)	(4,9)	(4,9,10)	(4,10)			
Н-5	2.01 m	2.07 d	2.13 dd	2.13 dd	2.43 dd			
		(5,7)	(9,14)	(10,15)	(4,15)			
		1.98 m	1.96 d	1.88 dd	2.30 dd			
		-	(4,14)	(4,15)	(10,15)			
H- 7	2.32 d (15)	2.31 dt	2.27 d(14)	2.64 d (15)	3.19d(17)			
		(1.5,11)						
	2.22 d (15)	2.13 dd (1.5,14)	2.23 d (14)	2.34 d (15)	2.32 d (17)			
Н-9	1.86 d (13)	1.85 dd	1.91 d(14)	2.20 dd	2.25 dd			
-	,	(2,14)		(3,15)	(3,15)			
	1.70 dd	1.48 m	1.73 d	1.82 d(15)	1.53 d(15)			
	(2,13)		(2.5,13)	,				
H-11	1.54t(12)	1.57 d (13.5)	1.60 t (12)	1. 59t(12)	1.58 t (12)			
	1.47 ddd	1.48 m	1.51 ddd	1.44 dd	1.49 ddd			
	(2, 8, 12)		(2.5,8,13)	(3,8,12)	(3,9,12)			
H-12	1.11s	1.09 s	1.09 s	1.03 s	1.01s			
H-13	1.04 s	1.03 s	1.04 s	1.01s	1.00 s			
H- 14	0.76s	0.92 s	0.78s	0.81 s	0.84 s			
H-15	1.02 d (7)	1.00 d(7)	0.96d(7)	1.00 d	0.99 d (7)			
				(6.5)				
СОМе			2.03 s	2.04 s	2.06 s			
				2.02 s	2.03 s			
					2.00 s			
ОН	1.77 s	2.07 s	1.63 s					
	1.64 s	1.57 s						

TABLE 2. ¹H-nmr Data² for Compounds 3, 4, 16–18.

*Reported as chemical shift (\delta), multiplicity, coupling constant in Hz (in parentheses); solvent $CDCl_3$.

this metabolite, and no unambiguous conclusions could be made about the stereochemistry of the C-3 methyl in **19**.

We tentatively assign structure 22 to the other minor metabolite. This compound has the molecular formula $C_{15}H_{18}O_4$ and is optically active. Its ir spectrum shows the presence of hydroxyl (3456 cm⁻¹), cyclopentanone carbonyl (1742 cm⁻¹), and a cyclopentenone α,β -unsaturated carbonyl (1704, 1629 cm⁻¹). Because compound 22 forms monoacetyl derivative 23 upon acetylation, the fourth oxygen of the molecular formula of 22 must be present as an ether. The ¹H nmr spectrum of 22 shows three methyl singlets. There is a vinyl hydrogen (δ 5.90, d) that shows allylic coupling to a methylene hydrogen (δ 2.76), which is further coupled to two other hydrogens (δ 3.33, 3.07, AB of ABX). Decoupling experiments show the presence of a second ABX spin system (δ 4.30, 3.98, 2.84). These data are consistent with structure 22. Further evidence for the tentative structural assignment is based on the spectral properties of its acetyl derivative 23. In the ¹H-nmr spectrum of 23, acetoxyl-induced anisotropic shifts are observed for the carbinyl hydrogen (1.17 ppm), H-8 (0.17 ppm), the geminal



methyl hydrogens, and the methylene hydrogens at C-15. These observations are consistent with the structure proposed. Because only very small amounts of compound **22** were obtained, no further experiments were possible.

Qualitative antifungal bioassay of the arthroconidial metabolites 1, 2, and 3 against several *Ceratocystis* species (7) reveals that these metabolites display weak antifungal activity, insufficient to account for the antifungal activity observed in the crude neutral extract unless their effect is synergistic. Further studies are required to establish which metabolites are responsible for the total antifungal activity.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—Distilled H_2O was used to prepare growth media and was redistilled using an all-glass distillation apparatus. Difco bacto potato dextrose broth (PDB) or potato dextrose agar (PDA) and yeast extract were utilized for growth media. Still culture fermentation was performed using 2.5-liter Fernback flasks. Celite 545 (American Chemicals Ltd) was used as a filter aid.

Reagent grade solvents were distilled prior to use. Skellysolve B refers to Skelly Oil Company petroleum ether, bp 62–70°. Analytical grade Et_2O (ACS 288) from freshly opened cans was used without further purification. Anhydrous MgSO₄ or anhydrous Na₂SO₄ was used a drying agent.

Cc was carried out using Brinkman Si gel 60 (0.08 mm, 200 mesh). E. Merck Si gel 60 (0.04–0.063 mm, 230–400 mesh) was used for flash cc (25). Analytical tlc was carried out using Si gel G containing 1% Retman P-1 electronic phosphor (General Electric) or on aluminum-foil-supported Si gel 60 (E. Merck, 0.25 mm, F_{254}) plates. The chromatograms were examined under uv light (λ 254 or λ 350 nm). The developed plates were visualized by spraying with aqueous 20% H_2SO_4 containing 1% vanillin (reagent A) or by dipping in 5% phosphomolybdic acid in aqueous 5% H_2SO_4 containing a trace of ceric sulfate (reagent B), followed by careful charring on a hot plate. Cholesterol was used as a standard and R'_f values are reported for the R_f of each metabolite relative to the R_f of cholesterol, which was assigned the value of 1.00 in the reported solvent system.

Melting points were determined on a Fisher-Johns melting point apparatus and are uncorrected. Ft-ir spectra were obtained on a Nicolet 7199 F.T. interferometer. Uv spectra were recorded on a Unicam SP 1700 spectrometer coupled to a 7470A plotter. Optical rotations were determined on a Perkin-Elmer model 141 automatic polarimeter. High resolution electron impact mass spectra (hreims) were recorded on an AEI MS-50 mass spectrometer coupled to a data processing DS-50 computer system. Cims and low resolution electron impact mass spectra (lreims) were determined on an AEI MS-4 mass spectra coupled to a DS-9 computer system. Data are reported as *m/z* (rel. int.). Only diagnostically significant peaks or those greater than 10% of the base peak are reported.

¹H-nmr and ¹³C-nmr spectra were recorded on a Bruker WH-400 spectrometer coupled to an Aspect 2000 computer system. Chemical shifts expressed in δ units are reported in ppm downfield from the inter-

nal standard TMS. Coupling constants, J, are expressed in Hz, and the following abbreviations are used: m = multiplet, s = singlet, d = doublet, t = triplet, q = quartet, br = broad.

CULTURING OF THE ARTHROCONIDIAL FUNGUS.—The strain of arthroconidial fungus used in these studies was isolated by Drs. Y. Hiratsuka and A. Tsuneda, Northern Forest Research Centre, Canadian Forestry Service, Edmonton, as a contaminant on a culture of *C. ulmi*. The fungus, believed to be a haploid basidiomycete, has not yet been fully identified. A culture was deposited at the University of Alberta Microfungus Collection under accession number UAMH 4262.

The culture was maintained at 4° in agar slant tubes containing PDA and 0.2% yeast extract (PDAY). Inoculum was prepared by adding 2 ml of sterile H_2O to a slant culture and gently scratching the surface of the culture with a sterile needle to produce a mycelial suspension. The suspension was used to inoculate two PDAY plates. Culture growth occurred at 15 to 20° over 2 weeks; then a mycelial suspension, prepared as described above from the plate culture, was transferred to two 500-ml Erlenmeyer flasks containing 200 ml sterilized PDB containing 0.2% yeast extract (PDBY). This culture was maintained at 15 to 20° on a gyrotory shaker for 2 weeks. Still cultures (10 liters) were prepared by transferring 25 ml of inoculum into ten Fernbach flasks each containing 1 liter of sterile PDBY.

The fungus was harvested after growth for 1 month at 15 to 20°. The mycelium was separated from the broth by filtration through glass wool. The mycelial mat was dried in a fume hood for 10 days, pulverized, and continuously extracted in a Soxhlet extractor with E_2O followed by EtOAc. Each organic extract was washed, dried, and concentrated to give crude mycelial extracts (E_2O extract, 0.15 g/liter; EtOAc extract, 0.24 g/liter).

The broth was clarified by filtration through a filter aid, then concentrated to $\frac{1}{3}$ volume at reduced pressure. The concentrate was continuously extracted for 2 days in a liquid-liquid extractor with Et₂O, then EtOAc. The organic extracts were dried and concentrated to give crude broth extracts (Et₂O extract, 0.11 g/liter; EtOAc extract, 0.04 g/liter).

PRELIMINARY FRACTIONATION OF THE CRUDE EXTRACTS.—The Et_2O -soluble extracts (2.15 g) were dissolved in EtOAc and separated into neutral, strong acidic, and weak acidic compounds by standard acid-base extraction (neutral extract, 0.97 g; strong acid extract, 0.15 g; weak acid extract, 0.39 g).

SEPARATION AND ISOLATION OF THE NEUTRAL METABOLITES.—The neutral extract was separated by flash chromatography using CH_2Cl_2 -MeCN (3:2). The fractions were analyzed by tlc and like fractions combined to give fractions A–E. Fraction A (83 mg) yielded **19** and **16**; fraction B (17 mg) **1**, **2**, and **19**; fraction C (147 mg) **1**, **2**, and **22**; fraction D (20 mg) **1**, **2**, and cyclohumuladiol; fraction E (50 mg) **3** and an unidentified metabolite. The individual components of each fraction were isolated by a combination of cc and preparative tlc. A complete description of the isolation of each of these sesquiterpenes, including the identification of cyclohumuladiol, may be found in Amouzou (7).

ARTHROSPORONE [1].— $R'_f 0.50$ [Me₂CO-C₆H₆ (3:2)]; recrystallized from Skellysolve B/Et₂O as colorless needles: mp 139–141°; [α]D – 140.8° (c = 0.9, CHCl₃); λ max (MeOH) 280 nm (ϵ = 650); Ft-ir (CHCl₃ cast) 3440, 2951, 2866, 1731, 1381, 1360, 1275, 1189, 1015 cm⁻¹; ¹³C nmr (CDCl₃) 216.4, (s, C-4), 91.0 (s, C-6), 87.0 (s, C-8), 60.6 (d, C-3), 58.9 (t, C-5), 56.9 (t, C-7), 55.5 (d, C-1), 54.6 (s, C-2), 49.9 (t, C-9), 44.7 (t, C-11), 40.2 (s, C-10), 29.6 (q), 26.9 (q), 11.3 (q), 8.3 (q); cims (NH₃) m/z [M + NH₄]⁺ 270 (100), [M]⁺ 252 (40); hreims (252.1722 calcd for C₁₅H₂₄O₃) found 252.1723 (36), 235 (38), 234 (28), 219 (13), 216 (2), 206 (12), 192 (89), 191 (24), 177 (22), 163 (21), 125 (100), 102 (25), 95 (33), 83 (40).

ANHYDROARTHROSPORONE [2]. $-R'_f 0.74$ [Me₂CO-C₆H₆ (2:3)]; recrystallized from Skellysolve B/Et₂O: mp 118–119°; [α]D +62° (c = 2.0, CHCl₃); λ max (MeOH) 230 nm (ϵ = 13,700); Ft-ir (CHCl₃ cast) 3461, 1693, 1632, 1466, 1448, 1372, 1364, 872 cm⁻¹; ¹³C nmr (CDCl₃) 211.6 (s, C-4), 177.0 (s, C-6), 122.9 (d, C-5), 92.7 (s, C-8), 63.4 (d, C-3), 57.7 (d, C-1), 55.9 (t, C-7), 53.4 (s, C-2), 44.0 (t, C-9), 43.3 (s, C-10), 41.8 (t, C-11), 30.2 (q), 28.2 (q), 21.8 (q, C-14), 9.5 (q, C-15); cims (NH₃) m/z [M + NH₄]⁺ 252 (2), [M + H]⁺ 235 (100), 234 (20); hreims (234.1616 calcd for C₁₅H₂₂O₂) found 234.1617 (63), 219 (32), 216 (34), 201 (23), 173 (23), 123 (78), 122 (100), 111 (10), 95 (13), 55 (20).

ARTHROSPOROL [3].— $R'_f 0.39$ [Me₂CO-C₆H₆ (3:2)]; recrystallized from Et₂O: mp 163–164°; [α]D -29° (c = 2.0, CHCl₃), -62.1 (c = 1.0, MeOH); Ft-ir (CHCl₃ cast) 3376, 2951, 2933, 2456, 1380, 1372, 1088, 1012 cm⁻¹; ¹³C nmr (CDCl₃) 91.0 (s, C-6 or -8), 90.3 (s, C-8 or -6), 76.8 (d, C-4), 60.9 (d, C-3 or -1), 58.5 (t, C-5 or -7), 58.2 (t, C-7 or -5), 56.3 (s, C-2), 52.1 (d, C-1 or -3), 49.0 (t, C-11 or -9), 45.3 (t, C-9 or -11), 40.0 (s, C-10), 30.7 (q), 28.6 (q), 12.9 (q), 12.5 (q); cims (NH₃) m/z [M + NH₄]⁺ 272 (100), 254 (18), 237 (15); hreims (236.1776 calcd for C₁₅H₂₄O₂) found [M - H₂O]⁺ 236.1778 (11), 218 (100), 203 (31), 190 (8), 182 (38), 174 (33), 150 (54), 139 (32), 135 (33), 127 (32), 125 (16), 123 (117), 121 (15), 110 (25), 109 (38), 107 (34), 95 (31), 83 (54), 69 (38), 55 (65). 4-0-ACETYLARTHROSPOROL [**16**].— R'_f 0.70 [Me₂CO-C₆H₆ (2:7)]; recrystallized from Skellysolve B/Et₂O: mp 132–135°; [α]D –60.9° (c = 1.4, CHCl₃); Ft-ir (CHCl₃ cast) 3450, 3440, 1720, 1261, 1021 cm⁻¹; cims (NH₃) m/z [M + NH₄]⁺ 314 (100), 296 (18), 236 (2); hreims (278.1883 calcd for C₁₇H₂₆O₃) found 278.1875 (0.2), 263 (0.2), 236 (3.5), 218 (100), 203 (18), 200 (0.8) 160 (14), 139 (13), 136 (41), 109 (25), 55 (23).

ARTHROSPORODIONE [19].— $R'_f 0.74$ [Me₂CO-C₆H₆ (2:7)]; [α]D -80.1° (ϵ = 2.5, CHCl₃); λ max (MeOH) 242 nm (ϵ = 7712), λ max (MeOH + 0.1 N HCl) 242 nm (ϵ = 8142), λ max (MeOH + 0.1 N NaOH) 242 nm (ϵ = 10,594); Ft-ir (CHCl₃ cast) 3450, 1738, 1700, 1688, 1637, 1374, 1089 cm⁻¹; ¹³C nmr (CDCl₃) 218.8 (s, C-4), 205.8 (s, C-11), 185.0 (s, C-8), 145.1 (s, C-1), 75.1 (d, C-9), 54.3 (s, C-10), 51.0 (d, C-3 or -6), 49.9 (d, C-6 or -3), 42.1 (s, C-2), 41.7 (t, C-5 or -7), 31.7 (t, C-7 or -5), 23.6 (q), 20.2 (q), 18.5 (q), 9.3 (q); cims (NH₃) m/z [M + NH₄]⁺ 266 (100), 248 (18), 235 (14); hreims (248.1412 calcd for C₁₅H₂₀O₃) found 248.1417 (42), 233 (100), 230 (4), 215 (5), 192 (11), 177 (19), 173 (7), 131 (13), 91 (17).

COMPOUND **22**.— R'_f 0.23 [EtOAc-pentane (2:3)]; [α]D +28.9° (c=0.92, CHCl₃); λ max (MeOH) 229 nm (ϵ = 19,400); Ft-ir (CHCl₃ cast) 3456, 3070, 1740, 1704, 1629, 1080 cm⁻¹; cims (NH₃) m/z [M + NH₄]⁺ 280 (100), 263 (43), 262 (21); hreims (262.1205 calcd for C₁₅H₁₈O₄) found 262.1207 (100), 234 (3), 191 (37), 163 (12), 72 (2); ¹H nmr (CDCl₃-D₂O) δ 5.90 (1H, d, 2.0 Hz, H-5), 4.29 (1H, t, 9.6 Hz, H-15 α), 3.98 (1H, dd, 9.0, 15.5 Hz, H-15 β), 3.92 (1H, s, H-11), 3.33 (1H, t, 9.0 Hz, H-8), 3.07 (1H, dd, 9.0, 15.5 Hz, H-7 α), 2.84 (1H, dd, 3.0, 9.5 Hz, H-3), 2.76 (1H, ddd, 2.2, 9.0, 16.0 Hz, H-7 β), 1.25 (3H, s, 10-Me), 1.15 (6H, s, 10-Me, 2-Me).

8-0-ACETYLARTHROSPORONE [5].—Arthrosporone [1] (2.4 mg) was dissolved in NEt₃ (2 ml), and N,N-dimethylaminopyridine (DMAP, 1.5 mg) and Ac₂O (4 drops) were added. The mixture was stirred at room temperature for 3 days, then concentrated. The residue was partitioned between EtOAc (20 ml) and 5% aqueous HCl (5 ml). The organic extract was washed with H₂O, dried and concentrated. Purification by flash chromatography gave an analytically pure sample of compound 5: mp 120–122°; R_f 0.58 [EtOAc-pentane (1:2)]; $[\alpha]D - 37.6^\circ$ (c = 5.0, CHCl₃); Ft-ir (CHCl₃ cast) 3449, 1736, 1382, 1368, 1249, 1017 cm⁻¹; hreims (294.1824 calcd for C₁₇H₂₄O₃) found 294.1818 (1), 252 (1.3), 234 (32), 216 (3), 192 (100), 125 (92), 110 (31).

6,8-0,0-DIACETYLARTHROSPORONE [6].—Arthrosporone [1] (2 mg) and Ac₂O (5 drops) were mixed in CH₂Cl₂ (2 ml). The solution was cooled in an ice bath, and *p*-toluenesulfonic acid (few grains) was added. The mixture was allowed to warm to room temperature over a period of 60 min. The mixture was diluted with CH₂Cl₂ (10 ml), washed with aqueous 5% NaHCO₃ (2 × 2 ml) and brine (2 ml), dried, and concentrated. Purification of the residue by chromatography gave diacetate **6** as a waxy solid: R'_f 1.18 [EtOAc-pentane (1:3)] Ft-ir (CHCl₃ cast) 1739, 1383, 1369, 1249, 1227, 1029 cm⁻¹.

8-0-ACETYLANHYDROARTHROSPORONE [7].—Anhydroarthrosporone [2] (6 mg), DMAP (catalytic amount), Ac₂O (5 drops), and NEt₃ (2 ml) was allowed to stand at room temperature for 7 h. The solution was concentrated and the residue was partitioned between CH₂Cl₂ (20 ml) and 5% aqueous HCl (2 × 2 ml). The organic phase was dried, concentrated, and purified by chromatography to give compound 7 as a colorless liquid which slowly solidified over time: mp 76–77°; R'_f 1.00 [EtOAc-pentane (1:2)]; [α]D +78.0° (c = 6.4, CHCl₃); λ max (MeOH) 228 nm (ϵ = 8273); Ft-ir (CHCl₃ cast) 1728, 1706, 1638, 1241 cm⁻¹; hreims (276.1719 calcd for C₁₇H₂₄O₃) found 276.1724 (2), 234 (4), 216 (100), 201 (39), 173 (36), 122 (27).

REDUCTION OF 1.—Arthrosporone [1] (5 mg) was dissolved in MeOH (2 ml), and NaBH₄ (7 mg) was added. The mixture was stirred at room temperature for 1 h. Aqueous HOAc (50%, 1 ml) was added and the mixture stirred for 2 min. MeOH was removed at reduced pressure, H₂O (2 ml) added, and the aqueous solution extracted with EtOAc. The organic extract was washed with brine, dried, concentrated, and purified by chromatography. Two products were obtained: arthrosporol [3] and epiarthrosporol [4]. Compound 4: mp 150°, R'_f 0.63 [C₆H₆-Me₂CO (3:2)]; [α]D – 64.0° (c = 0.1, MeOH); Ft-ir (CHCl₃ cast) 3301, 2950, 2930, 1378, 1364, 1067, 1024 cm⁻¹; hreims (254.1875 calcd for C₁₅H₂₆O₃) found 254.1875 (3), 236 (20), 218 (8), 182 (100), 177 (4), 174 (21), 164 (7), 150 (39), 139 (42), 127 (18), 124 (4), 121 (22), 111 (5), 83 (14), 55 (27).

DEHYDRATION OF 1 TO 2.—Arthrosporone [1] (2 mg) was dissolved in dry C_6H_6 (3 ml) containing molecular sieves (3Å) and *p*-toluenesulfonic acid (catalytic amount). The mixture was stirred at room temperature for 4 h. The solution was decanted and diluted with Et₂O. The organic solution was washed with 10% NaHCO₃ (2 × 2 ml), dried, concentrated, and purified by chromatography to give anhydroarthrosporone [2], identical in all respects with the natural metabolite. CONVERSION OF **2** TO BICYCLOKETONE **9**.—A solution of anhydroarthrosporone [**2**] (4.1 mg) in dry C₆H₆ (2 ml) was flushed with N₂ for 10 min. NaH (excess) was added as a suspension in C₆H₆ to the solution. The mixture was stirred at room temperature under an N₂ atmosphere for 5 h. The mixture was diluted with C₆H₆, quenched with 10% HOAc, then stirred until the excess NaH dissolved. The organic layer was separated, washed with brine, dried, and concentrated. The crude residue was purified by chromatography to give compound **9** in quantitative yield: mp 55–56°; R'_f 1.29 [C₆H₆-Me₂CO (8:1)]; [α]D – 202° (c = 1.95, CHCl₃); λ max (MeOH) 223 nm (ϵ = 13,400); Ft-ir (CHCl₃ cast) 1738, 1702, 1625 cm⁻¹; ¹H nmr (CDCl₃) δ 5.92 (1H, q, 1.6 Hz, H-5), 2.58 (1H, ddd, 12.5, 8.5, 1.5 Hz, H-1), 2.20 (1H, ddd, 18.0, 2.4, 1.5 Hz, H-9 α), 2.08 (1H, q, 7.5 Hz, H-3), 2.05 (3H, d, 1.6 Hz, H-7), 2.04 (1H, dd, 18.0, 1 Hz, H-9 β), 1.67 (1H, ddd, 13.0, 8.5, 2.5 Hz, HO-11 α), 1.45 (3H, s, H-14), 1.35 (1H, br t, 12.5 Hz, H-11 β), 1.19 (3H, s, H-13), 1.08 (3H, d, 7.5 Hz, H-15), 1.04 (3H, s, H-12); hreims (234.1614 calcd for C₁₅H₂₂O₂) found 234.1623 (59), 219 (7), 150 (37), 124 (36), 123 (100), 122 (90), 112 (26), 97 (15), 95 (50).

OXIDATION OF ARTHROSPOROL [3].—Arthrosporol [3] (3 mg) in CH_2Cl_2 and pyridinium chlorochromate (PCC, 2.2 mg) was stirred at room temperature for 2 h. The suspension was filtered through a short column of florisil. The florisil column was washed with Et_2O , and the organic filtrates were combined and concentrated. Crystallization from Skellysolve B/ Et_2O gave arthrosporone [1], identical in all respects with the natural metabolite.

REDUCTION OF 8-0-ACETYLANHYDROARTHROSPORONE [7].—Compound 7 (5 mg) was dissolved in MeOH (1.5 ml). CeCl₃ (2.2 mg) and NaBH₄ (10 mg) were added and the mixture stirred at room temperature for 1 h. The mixture was worked up in the usual way and the crude reaction product purified by chromatography to give three products: compounds **11**, **12**, and **13**.

Compound **11**.— R'_{f} 0.83 [C₆H₆-Me₂CO (3:1)]; Ft-ir (CHCl₃ cast) 3400, 1732, 1676, 1242, 1017 cm⁻¹; cims (NH₃) m/z [M + NH₄]⁺ 296 (6), 278 (6), 201 (100); hreims (278.1875 calcd for C₁₇H₂₆O₃) found 278.1873 (1), 218 (100), 203 (35), 200 (4), 106 (85), 91 (20).

Compound **12**.— $R'_f 0.93$ [C₆H₆-Me₂CO (3:1)]; Ft-ir (CHCl₃ cast) 3464, 1732, 1724, 1367, 1245, 1016 cm⁻¹; cims (NH₃) m/z [M + NH₄]⁺ 298 (64); hreims 220 (32), 202 (5), 162 (22), 161 (56), 148 (65), 110 (100), 107 (26).

Compound **13**.— R'_f 0.65 [C₆H₆-Me₂CO (3:1)]; Ft-ir (CHCl₃ cast) 3400, 1732, 1713, 1383, 1366, 1243, 1087, 1016 cm⁻¹; hreims 238 (0.5), 220 (41), 202 (66), 187 (23), 176 (16), 161 (52), 148 (100), 111 (19), 108 (34), 93 (25).

CORRELATION OF ANHYDROARTHROSPRONE [2] WITH COMPOUND 11.—A mixture of compound 2 (10 mg), 10% methanolic HOAc (4 drops), and activated Pd on charcoal (catalytic amount) in MeOH (2 ml) was stirred under an H₂ atmosphere for 30 min. The mixture was filtered through celite and the celite washed with MeOH. The combined filtrate was concentrated to yield ketone 15 in quantitative yield: mp 115–116°; R'_f 0.82 [C₆H₆-Me₂CO (7:2)]; Ft-ir (CHCl₃ cast) 3502, 1727 cm⁻¹; hreims (236.1772 calcd for C₁₅H₂₄O₃) found 236.1773 (16), 221 (9), 218 (20), 203 (9), 176 (71), 161 (33), 124 (39), 110 (100), 109 (65), 83 (5).

Compound **15** (8 mg) in MeOH (3 ml) and excess NaBH₄ was stirred for 1 h. HOAc (5 drops) was added and the mixture stirred for 5 min. Solvents were removed and the residue was partitioned between H₂O (5 ml) and EtOAc (3 × 10 ml). The organic extracts were dried and concentrated. Purification by chromatography gave two diols: major mp 139–140°; R'_f 0.14 [C₆H₆-Me₂CO (3:2)]; Ft-ir (CHCl₃ cast) 3347, 2953, 2932, 2869, 1040 cm⁻¹; minor R'_f 0.48 [C₆H₆-Me₂CO (4:1)].

Compound 12 (5 mg) was dissolved in 10% ethanolic KOH (3 ml), and the mixture was stirred at 80° for 20 min. The mixture was cooled to 0° and the reaction quenched with 20% HCl. Most of the EtOH was removed; the residue was diluted with H₂O and extracted with EtOAc. The organic extract was dried, concentrated, then purified by chromatography to give a single diol which was recrystallized from $CH_2Cl_2/Skellysolve B: mp 138-140°$; $R'_f 0.22 [C_6H_6-Me_2CO (1:4)]$; Ft-ir (CHCl₃ cast) 3345, 2952, 2933, 2869, 1045 cm⁻¹; cims (NH₃) m/z [M + NH₄]⁺ 238 (64); ms 220 (13), 205 (6), 202 (50), 179 (32), 166 (56), 148 (100), 120 (27), 111 (55), 109 (69), 108 (96), 95 (52), 55 (42).

ACETYLATION OF ARTHROSPOROL: 4-0-ACETYLARTHROSPOROL [16].—A solution of 3 (3.4 mg), Ac_2O (5 drops), and C_5H_5N (1.5 ml) was stirred at room temperature for 24 h. Workup in the usual way and purification by chromatography gave compound 16 which was recrystallized from Skellysolve B/ Et_2O (mp 150°).

4,8-0,0-DIACETYLARTHROSPOROL [17].—Arthrosporol [3] (4.7 mg), DMAP (catalytic amount), and Ac_2O (5 drops) were mixed in NEt₃ and allowed to stand at room temperature for 3 days. The mixture was worked up in the usual way. Purification of the crude acetylate by chromatography gave compound 17 as a waxy solid: $R'_f 0.86$ [C₆H₆-Me₂CO (2:7)]; Ft-ir (CHCl₃ cast) 3512, 2952, 1732, 1717, 1249, 1020 cm⁻¹; Ireims 278 (3), 219 (50), 218 (100), 203 (13), 190 (17), 109 (30), 95 (17), 55 (14), 45 (58).

4,6,8-0,0,0-TRIACETYLARTHROSPOROL [18].—Arthrosporol [3] (2.4 mg) and p-toluenesulfonic acid were dissolved in Ac₂O (1 ml), and the solution was stirred at room temperature for 3 h. The solution was diluted with Et_2O (10 ml), washed (5% aqueous NaHCO₃, H₂O, brine), dried, and the solvent removed. The residue was chromatographed to give pure 18 as a viscous oil: R'_f 1.43 [C₆H₆-Me₂CO (1:8)]; Ft-ir (CHCl₃ cast) 1738, 1249, 1225, 1020 cm⁻¹; cims (NH₃) m/z [M + NH₄]⁺ 398 (100); hreims 321 (1), 260 (11), 218 (8), 200 (100).

ACETYLDEHYDROARTHROSPORODIONE [20].—Compound 19 (4 mg), C_5H_5N (1.5 ml), and Ac_2O (4 drops) were stirred at room temperature for 12 h. The reaction mixture was worked up in the usual way. Purification by preparative tlc [$Me_2CO-C_6H_6$ (1:19)] gave pure 20: R'_f 1.07 [$C_6H_6-Me_2CO$ (1:8)]; [α]D – 13.8° (c = 1.6, CHCl₃); Ft-ir (CHCl₃ cast) 1740, 1710, 1632, 1328, 1374, 1232 cm⁻¹; ¹H nmr (CDCl₃) δ 5.60 (1H, t, 2.5 Hz, H-9), 2.97 (1H, m, H-6), 2.87 (1H, ddd, 16, 9, 2.4 Hz, H-7 α), 2.59 (1H, dd, 20, 9 Hz, H-9), 2.42 (1H, ddd, 19, 4, 1.5 Hz, H-5 β), 2.40 (1H, qd, 7, 1.5 Hz, H-3), 2.25 (1H, ddd, 16, 7, 2.5 Hz, H-7 β), 2.17 (3H, s, CH₃CO), 1.26 (3H, s, Me), 1.1 (3H, s, Me), 1.08 (3H, d, 7 Hz, H-15), 1.00 (3H, s, Me); ¹³C nmr (CDCl₃) δ 217.5 (s, C-4), 204.9 (s, C-11), 181.4 (s, C-3), 170.3 (s, MeCO), 147.5 (s, C-1), 75.5 (d, C-9), 53.9 (s, C-10), 51.0 (d, C-3), 49.9 (d, C-6), 42.1 (s, C-2), 41.7 (t, C-5), 31.7 (t, C-7), 24.1 (q), 20.7 (q), 19.9 (q, MeCO), 18.6 (q), 9.4 (q); cims (NH₃) m/z [2M + NH₄]⁺ 598 (0.3), [M + NH₄]⁺ 308 (100); hreims (290.1512 calcd for C₁₇H₂₂O₄) found 290.1524 (7), 248 (89), 233 (100), 177 (25), 174 (119), 91 (19).

OXIDATION OF DEHYDROARTHROSPORODIONE.—Compound **19** (2.7 mg) was dissolved in CH_2CI_2 , and PCC (2.3 mg) was added. The yellow suspension was stirred at room temperature for 20 h. The suspension was filtered through florosil and the adsorbent washed with CHCI₃. The filtrate and washings were combined and concentrated. Purification of the residue by chromatography gave triketone **21** as a low melting solid: $R'_f 1.29 [C_6H_6-Me_2CO (1:8)]; [\alpha]D - 153.9^\circ (c = 0.7, CHCI_3); \lambda max (MeOH) 247$ nm ($\epsilon = 4762$); Ft-ir (CHCI₃ cast) 1736, 1696, 1616 cm⁻¹; ¹H nmr (CDCI₃) δ 3.10 (1H, dd, 17, 9 Hz, H-7 α), 3.07 (1H, m, H-6), 2.74 (1H, dd, 18.5, 9 Hz, H-5 α), 2.51 (1H, dd, 17, 6 Hz, H-7 β), 2.39 (1H, m, H-3), 2.21 (1H, dd, 18.5, 5 Hz, H-5 β), 1.23 (3H, s, Me), 1.19 (3H, s, Me), 1.16 (3H, s, Me), 1.15 (3H, d, 7 Hz, H-15); hreims (246.1251 calcd for C₁₅H₁₈O₃) found 246.1257 (100), 231 (7), 218 (3), 190 (29), 189 (53), 176 (50).

ACETYLATION OF COMPOUND 22.—Compound 22 (1 mg), C_5H_5N (1 ml), and Ac_2O (2 drops) were stirred at room temperature for 24 h at which time the starting material was recovered unchanged. The solution was concentrated to dryness and the residue dissolved in NEt₃ (1 ml). Ac_2O (2 drops) and DMAP (ca. 0.5 mg) were added and the mixture stirred overnight. Workup in the usual way gave a complex mixture (tlc). Purification by chromatography gave a small amount of a uv-active compound (compound 23): Ft-ir (CHCl₃ cast) 1744, 1709, 1632, 1233 cm⁻¹; ¹H nmr (CDCl₃) δ 5.88 (1H, d, 2 Hz, H-5), 5.09 (1H, s, H-11), 3.97 (1H, dd, 7, 4 Hz, H-15 α), 3.52 (1H, t, 8 Hz, H-8), 3.51 (1H, t, H-15 β), 3.03 (1H, dd, 15, 9 Hz, H-7 α), 2.84 (1H, ddd, 15, 8.5, 1.5 Hz, H-7 β), 2.78 (1H, dd, 7, 4 Hz), 2.20 (3H, s, CH₃CO), 1.32 (3H, s, H-13), 1.27 (3H, s, H-12), 1.14 (3H, s, H-14).

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